

ARTICLE

Characterization of Pediocin PaF-11 from *Pediococcus acidilactici* F-11 as Biopreservatives

Tri Marwati^{1,*}, Nur Richana², Eni Harmayani³, and Endang S. Rahayu³

1 Assessment Institute for Agricultural Technology of Yogyakarta, Jl. Stadion no 22 Maguwoharjo, Wedomartani, Ngemplak, Sleman, Yogyakarta, Indonesia

2 Indonesian Center for Agricultural Postharvest Research and Development, Jl. Tentara Pelajar No 12, Cimanggu, Bogor, Indonesia

3 Faculty of Agricultural Technology, Gadjah Mada University, Yogyakarta, Indonesia, Jl. Flora No.1, Bulaksumur, Sleman, Yogyakarta, Indonesia

E-mail: watipasca@yahoo.com

Pediocin PaF-11 is bacteriocin from *Pediococcus acidilactici* F-11. The aim of this study was to know the characteristics of pediocin PaF-11 as food biopreservatives. This study used *P. acidilactici* F-11 and *Lactobacillus pentosus* LB 42 as the pediocin producing and indicator strain, respectively. Both strains were obtained from the Food Nutrition Culture Collection, Gadjah Mada University. The *P. acidilactici* F-11 was grown to produce the pediocin in TGE liquid medium at a pH of 6.5 and an incubation temperature of 37°C for 18 hours. The pediocin PaF-11 was extracted and purified according to the adsorption-desorption methods and its activity was determined by the well-inhibition methods. The stability of pediocin against temperature and storage condition was characterized. The purified pediocin PaF-11 was stable after heating at a temperature of 100°C for 30 minutes or temperature 121° C for 15 minutes and storage at a temperature of 30°C and 4°C for 11 and 13 weeks. It was also active over a wide range of pH values from 3 to 8. The stability of pediocin PAF-11 to temperature is also supported by the following amino acid sequence results. Amino acid from the N-terminal of pediocin PAF-11 was sequenced: MKKIEKLTEKEMANIIGGKYYGNGVTCGKHSCSVDWKGKATTCIINNGAMAWATGGHGHQGNHKC). The pediocin PaF-11 is in the consensus sequence YGNGVXCXXXCVXXXXA which indicates that pediocin PaF-11 is belong to class IIa bacteriocin. These characteristics make pediocin PaF-11 suitable as a food preservative, especially products that involve processing on pH such as in the process of fermentation and processing at high temperatures as well as storage.

Introduction

One of the antibacterial substances that is suitable for food preservation is bacteriocin produced by lactic acid bacteria. Bacteriocin is a peptide or peptide complexes are synthesized in ribosomes. nisin produced by *Lactococcus lactis* subsp. *lactis* is the only commercial product that has a status of GRAS (Generally Recognized as Safe) and accepted by the USA FDA (Food and Drug Administration) as a food preservative. Nisin has been applied in most of the major food-producing country. However, nisin are less effective against gram-negative bacteria, yeast and fungi and solubility as well as its stability decreased significantly on the conditions of the meat which is neutral and alkaline.

Pediocin of *Pediococcus* draws attention to researched because some researchers have proved that the pediocin of *P. acidilactici*, *P. damnosus*, *P. parvulus* and *P. pentosaceus* has the ability to inhibit pathogens and spoilage bacteria so that it could potentially be used as a food preservative. The potential is supported by the benefits that can improve the weaknesses of nisin, among others in its stability in a wide range of pH (Huang et al., 2009; Anastasiadou et al., 2008a; Todorov and

Dicks, 2009; Osmanagaoglu, et al., 2011) as well as heat treatment (Anastasiadou, et al., 2008b).

On the assumption that a specific bacteriocin will have its own unique properties and usefulness in targeting microbial pathogens, isolation and purification of new bacteriocins will always prove beneficial. In-depth characterization of the purified form of new isolates will provide information for economically viable mass production and downstream processing (Coventry et al., 1996), together with useful data for making manipulations to improve bacteriocin properties and effectiveness (De Vuyst and Vandamme, 1994), and to hasten consumer acceptability of bacteriocintreated products.

The pediocin PaF-11 produced by *Pediococcus acidilactici* F-11 has potential use as biological food preservatives. In order to use it in the most effective way as biological preservative, the most important factors to be considered are stability at various pH, temperature and storage condition of pediocin PaF-11. To support the application of pediocin PaF-11, research that aims to know sequence of amino acids and the molecular weight were also needed.

Materials and Methods

Bacterial cultures and media

P. acidilactici F-11 and *Lactobacillus pentosus* LB42 were used as pediocin PaF-11 producer and indicator bacteria respectively, both of which were obtained from the Food and Nutrition Culture Collection (FNCC), Center for Food and Nutrition Studies of Gadjah Mada University in Yogyakarta. *P. acidilactici* F-11 produced pediocin PaF-11 in the TGE broth consist of tryptone (1%), glucose (1%), yeast extract (1%), tween 80 (0.2%), MnSO₄.H₂O (0,005%), and MgSO₄.7H₂O (0,0056%) at pH 6.5 within 18 hours incubation at 37 °C. The assay plate had a bottom layer of TGE agar (TGE broth plus 1.5% agar) and a top layer of TGE soft agar (TGE broth plus 0.75% agar)(Biswas et al., 1991).

Production and purification of pediocin PaF-11

Partial purification of the pediocin PaF-11 was performed by the adsorption desorption methods (Yang et al., 1992) was applied. Fresh culture of *P. acidilactici* F11 was inoculated at a rate of 0.1% in modified TGE broth at pH 6.5 and pre-sterilized at 15 psi for 15 min (Biswas et al., 1991). *P. acidilactici* F11 was grown 18 hour in TGE broth. Broth culture was heated at 100°C for 30 minutes to kill the cells after its pH adjusted to 6.5. The mixture was kept at 4 °C for 24 hour by mixing with stirrer. The cells were harvested by centrifugation at 15,000 rpm for 15 minutes. After the cells had been washed with 5mM sodium phosphate (pH 6.5), the cells were then resuspended in H₂O, and 0.1 M NaCl was added and the pH of the mixture was adjusted to 2.0. The mixture was kept at 4°C for 24 hr by mixing with stirrer. Cell suspensions were then centrifuged at 29,000 rpm for 20 minutes, and the supernatants were freeze-dried.

Antimicrobial activity assay of pediocin PaF-11

Pediocin PaF-11 activity was determined by well diffusion agar methods. The assay plate had a bottom layer of TGE agar (TGE broth plus 1.5% agar) and a top layer of TGE soft agar (TGE broth plus 0.75% agar) and was seeded with about 10⁶ *L. pentosus* LB42 cells. Then, 5mm wells were bored out and each was filled with 20µl of the pediocin PaF-11 fractions which were serially diluted. The agar plates were incubated at 37°C overnight. The highest dilution that produced a clear zone was multiplied by 50 (1ml/20µl) to obtain the activity units per milliliter (Biswas et al., 1991; Kim et al., 1993).

Effect of heat treatment on antimicrobial activity of pediocin PaF-11

Analysis of the effect of heat treatment on antimicrobial activity of pediocin PaF-11 was done by heating the pediocin PaF-11 at 100°C for 15 and 30 minutes (Huang et al., 2009) and autoclaving at 121°C for 5 and 15 minutes (Schneider et al., 2006; Huang et al., 2009). All

samples were cooled and assayed for activity (Elegado et al., 1997; Osmanagaoglu et al., 1998).

Effect of pH on antimicrobial activity of pediocin PaF-11

Pediocin PaF-11 was adjusted with sterile 10 mM/l NaOH or 10 mM/l HCl to different pH levels between 3 (Osmanagaoglu et al., 2001) to 10. Samples were maintained at 37 °C for 2 hours (Huang et al., 2009). The samples were then adjusted to pH 7.0 with sterile 4 mM/l phosphate buffer and assayed for activity (Elegado et al., 1997; Osmanagaoglu et al., 1998).

Effect of storage condition on antimicrobial activity of pediocin PaF-11

For storage stability determination, pediocin was storage in the cool room at 4°C and room with temperature 25-30°C (Wu et al., 2004; Mathys et al., 2007; Anastasiadou, et al., 2008a). Both sampels were assayed for activity every week periodically.

Amino acid sequencing of pediocin PaF-11

Plasmid isolation of *P. acidilactici* F-11 with alkaline lysis done by Sambrook et al., (1989) and Kalmokoff et al., (2003) methods. Plasmid DNA of *P. acidilactici* F-11 isolated then used as templates for the gene pediocin (papA) amplification through the PCR analysis (Huang et al., 2009; Todorov and Dick, 2009). The Primer is designed based on the sequence of papA gene in the GenBank database uses the program primer3 online (<http://frodo.wi.mit.edu/primer3/>), there were: primer 1 (Forward) 5'-GCGCGTATTAAGGATAATTT-3' and primer 2 (Reverse) : 5'-TTTATTGATGCCAGCTCAGC-3'. Nucleotide sequencing of DNA is done by DNA sequenser. Molecular weight pediocin PaF-11 performed by calculation used to use a program that refers to the EnCor Biotechnology Inc. © 2011 Analysis.

Results and Discussion

Stability of pediocin PaF-11 on heat treatment

Heat treatment at 100 °C for 15 and 30 minutes and autoclaving at 121 °C for 5 and 15 minutes did not destroy the activity of pediocin PaF-11 activity. Antibacterial activity of pediocin PaF-11 after heating at 100°C and autoclaving at 121°C using *L. pentosus* LB42 as indicator seen in Table 1.

Table 1. Antibacterial activity of pediocin PaF-11 after heat treatment using *L. pentosus* LB42 as indicator

Heat treatment	Pediocin PaF-11 activity (AU/ml)
100 °C for 15 min	1500
100 °C for 30 min	1500
121 °C for 5 min	1500
121 °C for 15 min	1500

Previous research showed that pediocin PaF-11 resistant to heat. It have been reported that bacteriocin from *P. acidilactici* strain Hansen remain stable after heating at 115°C for 15 min (Lozano et al., 2002) *P. acidilactici* SJ-1 at 65-121°C (Schved et al., 1993), *P. acidilactici* M; *P. acidilactici* F; *P. acidilactici* C20; and *P. acidilactici* PO2 at 121 °C for 15 min (Elegado et al., 1997; Osmanaglaogu et al., 1998; Halami et al., 2000; and Coventry et al., 1995). The recently isolated and characterized pediocin SA-1 from *P. acidilactici* NRRL B5627 was also found to be heat stable for up to 60 min at 121°C (Papagianni and Anastasiadou, 2009).

The thermotolerance feature of pediocin PaF-11 might be related to the molecular structure of the bacteriocin, usually composed by small peptides without tertiary structure (Parada et al., 2007) and has low molecular weight (Osmanaglaogu et al., 2001; Papagianni dan Anastasiadou, 2009). Another possibility is that because pediocin PaF-11 contain cysteine (Nugroho and Rahayu, 2003). These characteristics make pediocin PaF-11 suitable as a food preservative, especially for developing products that involve process at high temperature such as pasteurization.

Stability of pediocin PaF-11 on pH treatment

Pediocin PaF-11 was found to be stable over a wide pH range between 3 and 8 (Table 2). Activity was lost at pH 9, the loss of activity at higher pH could be due to degradation of the molecule (Osmanaglaogu et al., 1998). This is similar to the properties reported for bacteriocins produced by other lactic acid bacteria such as bacteriocin from *P. acidilactici* SJ-1 and *P. acidilactici* F stabil at pH 3-9 (Schved et al., 1993; Osmanaglaogu et al., 1998); *P. acidilactici* M stabil at pH 1-12 (Elegado et al., 1997), *P. acidilactici* NRRL B5627 stabil at pH 3-12 (Anastasiadou et al., 2008), *P. pentosaceus* ST18 stabil at pH 2-12 (Todorov dan Dicks, 2005) *P. pentosaceus* 05-10 stabil at pH 2-8 (Huang et al., 2009) dan *P. pentosaceus* Pep1 stabil at pH 3-8 (Osmanaglaogu et al., 2001).

Table 2. Antibacterial activity of pediocin PaF-11 after pH treatment using *L. pentosus* LB42 as indicator

pH	Pediocin PaF-11 activity (AU/ml)
3	1500
4	1500
5	1500
6	1500
7	1500
8	1200
9	0
10	0

The increased antibacterial activity observed at low pH may be the result of aggregation of hydrophilic peptides is less likely to occur, and, thus, more molecules should be available to interact with sensitive cells (Jack et al., 2005). Stability of pediocin PaF-11 at a wide range of pH (between 3 and 8) make it suitable as a food

preservative, especially for developing products that involve process wherein pH plays important role such as fermentation

Stability of pediocin PaF-11 on storage treatment

Activity of pediocin PaF-11 remained unaffected after 11 weeks storage at 4°C, and decreased after 11 weeks. Total loss of bacteriocin activity was already observed after 11 weeks storage at 30°C (Table 3). The loss of activity of pediocin during storage in the presence of air could be due to oxidation of methionine. In liquid preparation, aggregation of the molecules can also produce an apparent loss (Naidu, 2000).

Similar result, pediocin SA-1 from *P. acidilactici* NRRL B5627 was also found to be stable during storage. Storage for 4 weeks at -80, -20, 4 and 30°C did not affect its antimicrobial activity. This was not impaired even following incubation at 30°C for 1 week at pH values ranging between 3.0 to 12.0 (Papagianni and Anastasiadou, 2009). Activity of bacteriocin from *P. acidilactici* UVA1 unaffected after one month storage at 4°C at pH values from 2 to 8. At pH 10 and 11, total loss of activity was already observed after 1 week storage and only 60% of the initial activity was still measurable after 1 month at pH 9 (Mathys et al., 2007).

Table 3. Antibacterial activity (b) of pediocin PaF-11 after storage at 4 °C and 30 °C using *L. pentosus* LB42 as indicator

Storage time (Week)	Pediocin PaF11 activity (AU/ml)	
	Storage temperature (4°C)	Storage temperature (30°C)
1	1500	1500
2	1500	1500
3	1500	1500
4	1500	1500
5	1500	1250
6	1500	1500
7	1500	1500
8	1500	1500
9	1500	500
10	1500	500
11	1500	250
12	1500	0
13	1000	0

Amino acid sequence of pediocin PaF-11

The amino acid from the N-terminal of pediocin PAF-11 was sequenced : Methionine-Lysine-Lysine-Isoleucine-Glutamate-Lysine-Leusine-Threonine-Glutamate-Lysine-Glutamate-Methionine-Alanine-Asparagine-Isoleucine-Isoleucine-Glycine-Glycine-Lysine-Tyrosine-Tyrosine-Glycine-Asparagine-Glycine-Valine-Threonine-Cysteine-Glycine-Lysine-Histidine-Serine-Cysteine-Serine-Valine-Aspartate-Tryptophan-Glycine-Lysine-Alanine-Threonine-Threonine-Cysteine-Isoleucine-Isoleucine-Asparagine-Asparagine-Glycine-Alanine-Methionine-Alanine-Tryptophan-Alanine-Threonine-Glycine-Glycine-Histidine-Glutamine-Glycine-Asparagine-Histidine-Lysine-Cysteine

(MKKIEKLTEKEMANIIGGKYYGNGVTCGKHSCSVDWG KATTCIINNGAMAWATGGHQGNHKC). Based on pediocin PaF-11 sequence ieMKKIEKLTEKEMANIIGGKYYGNGVTCGKHSCSVDWG KATTCIINNGAMAWATGGHQGNHKC, and calculation using a program that refers to the EnCor Biotechnology Inc. © 2011 Analysis, the pediocin PaF-11 has a molecular weight of around 6.65. kDa.

Table 4. Nuclotide sequence of encoding gen and amino acid sequence of pediocin PaF-11

DNA Amino Acid Symbol	ATG AAA AAA ATT GAA AAA ITA ACT GAA AAA GAA ATG GCC AAT ATC ATT Met Lys Lys K Ile Glu Lys Lys Glu Met Ala Asn Ile Ile I
DNA Amino Acid Symbol	GGT GGT AAA TAC TAC TGG GGT AAT GGG GTT ACT IGT GGC AAA CAA ICC IGC Gly Gly Lys K Tyr Y G N G Val Thr Cys Gly Lys His Ser Cys C
DNA Amino Acid Symbol	TCT GTT CAC TGG GGT AAG GCT ACC ACT TGC ATA ATC AAT AAT GGA GCT Ser Val Asp D W G Lys Ala Thr Thr Cys Iso Iso Asn Asn Gly Ala A
DNA Amino Acid Symbol	ATG GCA TGG GCT ACT GGT GGA CAA GGT AAT CAT AAA TGC Met Ala W A T G Gly Gly His Gln H G N H Lys Cys C

Remarks : Class II a bacteriocin consensus : **KYYGNGVXCXXXXCXVXXXXA**
 Hidrofobic amino acid : metionin, isoleusin, leusin, alanin, tirosin, valin, dan triptofan

The pediocin PaF-11 is in the consensus sequence **YYGNGVXCXXXXCXVXXXXA** which indicates that pediocin PaF-11 is belong to class IIa bacteriocin. The sequence of pediocin PaF-11 has a high homology (100%) with pediocin PA-1 (from *P. acidilactici* PAC 1.0 (Marugg et al., 1992) and *P. pentosaceus* IE-3); prepediocin PA-1 (from *P. acidilactici* K10) (Moon dan Kim, 2004); pediocin Ach (from *P. acidilactici* H Motlagh, et al., 1992; *P. acidilactici* LB42-923 (Motlagh et al.,

1994); *P. parvulus* ATO77 (Miller et al., 2005), and *P. pentosaceus* S34 Miller et al., 2005) and prepediocin CP2 from *P. acidilactici* MTCC 5101 (Balgir et al., 2010), and pediocin from *P. acidilactici* CFR K7 (Halami et al 2005) and *P. acidilactici* PED 01 (Laoye and Dodd, 2008). Based on pediocin PaF-11 sequence and calculation using a program that refers to the EnCor Biotechnology Inc. © 2011 Analysis, the pediocin PaF-11 has a molecular weight of around 6.65. kDa. That result showed that pediocin PaF-11 has low molecular weight, is similar to pediocins isolated by various *Pediococcus*, ie the pediocin Ach (from *P. acidilactici* PAC 1.0; *P. acidilactici* LB42-923; *P. parvulus* ATO77) and pediocin PA-1 (from *P. acidilactici* K10; *P. pentosaceus* IE-3; *P. acidilactici* MTCC 5101) with molecular weight around 6644 Da. The pediocin PaF-11 is a larger molecule compared to the 3660, 3500, 5370, 5000. 6500, dan 4623 Da molecule of pediocin SA-1, PA-1, SM-1, K23-2, 05-10, and Ach. These pediocin PaF-11 characteristics make pediocin PaF-11 suitable as a food preservative, especially products that involve processing at high temperatures because protein with low molecular weight has stability to heat process. The molecular weight of pediocin PaF-11 showed that it resembles known pediocins and belongs therefore to the class IIa of bacteriocins by according to the classification Drider et al., (2006); Aly et al., (2006) dan Nissen-Meyer dan Nes (1997).

Conclusions

Pediocin PaF-11 was stable at pH of 3-8 and autoclaving (121°C for 15 min). Pediocin PaF-11 was stable during 11 and 13 weeks storage at 30°C and 4°C respectively. The pediocin PaF-11 is in the consensus sequence **YYGNGVXCXXXXCXVXXXXA** which indicates that pediocin PaF-11 is belong to class IIa bacteriocin. Based on that sequence, the pediocin PaF-11 has a molecular weight of around 6.65. kDa. These pediocin PaF-11 characteristics make pediocin PaF-11 suitable as a food preservative, especially products that involve processing at high temperatures because protein with low molecular weight has stability to heat process.

Conflict of Interest

All the authors declare that they have no conflict of interest.

References

- 1 Aly S, Ouattara CAT, Bassole IHN, Traore SA. 2006. Bacteriocins and lactic acid bacteria - a minireview. African Journal of Biotechnology 5(9): 678-683.
- 2 Anastasiadou S, Papagianni M, Filiouis G, Ambrosiadis I, Koidis P. 2008a. Growth and metabolism of a meat isolated strain of *Pediococcus pentosaceus* in submerged fermentation. Purification, characterization and properties of the produced pediocin SM-1. Enzyme Microbiology Technology 43:448-454.

- 3 Anastasiadou S, Papagianni M, Filiouis G, Ambrosiadis I, Koidis P. 2008b. Pediocin SA-1, an antimicrobial peptide from *Pediococcus acidilactici* NRRL B5627: Production conditions, purification and characterization. *Bioresource Technology* 99:5384–5390.
- 4 Anastasiadou S., M. Papagianni, G. Filiouis, I. Ambrosiadis and P. Koidis. 2008b. Pediocin SA-1, an antimicrobial peptide from *Pediococcus acidilactici* NRRL B5627: Production conditions, purification and characterization. *Bioresource Technology* 99:5384–5390.
- 5 Balgir P, Bhatia P. Kaur B. 2010. Sequence analysis and homology-based modeling to assess structure-activity relationship of pediocin CP2 of *Pediococcus acidilactici* MTCC 5101. *Indian Journal of Biotechnology* 9:431-434.
- 6 Biswas SR, Ray P, Johnson MC, Ray B. 1991. Influence of growth condition on the production of bacteriocin, pediocin AcH, by *Pediococcus acidilactici* H. *Appl. Environ. Microbiol.* 57:1265-1267.
- 7 Coventry MJ, Muirhead K, Hickey MW. 1995. Partial characterisation of pediocin PO, and comparison with nisin for biopreservation of meat products. *International Journal of Food Microbiology* 26: 133-145.
- 8 Coventry MJ, Gordon JB, Alexander M, Hickey MW, Wan J. 1996. A food-grade process for isolation and partial purification of bacteriocins of lactic acid bacteria that uses diatomite calcium silicate. *Appl. Environ. Microbiol.* 62. 1764- 1769.
- 9 De Vuyst, L. Vandamme, E.J. 1994. Lactic acid bacteria and bacteriocins: their practical importance. In: De Vuyst, L., Vandamme, E.J. *ŽEds., Acteriocins of Lactic Acid Bacteria: Microbiology, Genetics and Applications.* Blackie Academic and Professional, London. pp. 1–11. diatomite calcium silicate. *Appl. Environ. Microbiol.* 62. 1764- 1769.
- 10 Drider D, Fimland G, Hechard Y, Mc Mullen, Prevost H. 2006. The continuing story of class IIa Bacteriocins. *Microbiology and Molecular Biology Reviews.* 70(2):564-582.
- 11 Elegado F., Kim WJ, Kwon DY. 1997. Rapid purification, partial characterization, and antimicrobial spectrum of the bacteriocin, pediocin AcM, from *Pediococcus acidilactici* M. *International Journal of Food Microbiology* 37:1-11.
- 12 Halami PM, Ramesh A, Chandrashekar A. 2000. Megaplasmid encoding novel sugar utilizing phenotypes, pediocin production and immunity in *Pediococcus acidilactici* C20. *Food Microbiology* 17: 475-83.
- 13 Halami PM, Ramesh A, Chandrashekar A. 2005. Fermenting Cucumber, a Potential Source for the Isolation of Pediocin-Like Bacteriocin. *World Journal of Microbiology and Biotechnology.* 21:1351-1358.
- 14 Huang Y, Luo Y, Zhai Z, Zhang H, Yang, Ch, Tian H, Zheng Li, Feng J, Liu H, HaoY. 2009. Characterization and application of an anti-*Listeria* bacteriocin produced by *Pediococcus pentosaceus* 05-10 isolated from Sichuan Pickle, a traditionally fermented vegetable product from China. *Food Control.* 20(11): 1030-1035.
- 15 Jack RW, Tagg JR, Ray B. 1995. Bacteriocins of Gram Positive Bacteria. *Microbiology Reviews* 59:171-200.
- 16 Kalmokoff ML, Cyr TD, Hefford MA, Whitford MF, Teather RM. 2003. Butyriovibriocin AR10, a new cyclic bacteriocin produced by ruminal anaerobe *Butyriovibrio fibrisolvens* AR10: characterization of the gene and peptide. *Canadian Journal of Microbiology.* 49(12): 763-773.
- 17 Kim WJ, Hong. SS, Cha SK, Koo YJ. 1993. Use of bacteriocinogenic *Pediococcus acidilactici* in sausage fermentation. *J. Microbial. Biotechnol.* 3:199-203.
- 18 Lozano JCN, Reguera-Useros JI, Pela´ ez-Marti´ nez MC, Hardisson de la Torre A. 2002. Bacteriocinogenic activity from starter cultures used in Spanish meat industry. *Meat Science* 62 : 237–243.
- 19 Marugg JD, Gonzalez CF, Kunka BS Ledebøer AM, Pucci MJ, Toonen MY, Walker SA, Zoetmulder LC, Vandenbergh PA. 1992. Cloning, expression, and nucleotide sequence of genes involved in production of pediocin PA-1, and bacteriocin from *Pediococcus acidilactici* PAC1.0. *Applied and Environmental Microbiology.* 58(8):2360-2367.
- 20 Mathys S, Ueli von Ah, Lacroix C, Staub E, Mini R, Cereghetti T, Meile L. 2007. Detection of the pediocin gene *pedA* in strains from human faeces by real-time PCR and characterization of *Pediococcus acidilactici* UVA1. *BMC Biotechnology* 7:55 doi: 10.1186/1472-6750-7-55.
- 21 Miller KW, Ray P, Steinmetz T, Hanekamp T. Ray B. 2005. Gene organization and sequences of pediocin AcH/PA-1 production operons in *Pediococcus* and *Lactobacillus* plasmids. *Letters in Applied Microbiology* 40:56-62.
- 22 Moon GS, Kim WJ. 2004. Characterization of pediocin operon in *Pediococcus acidilactici* K10. Submitted (AUG-2004) to the EMBL/GenBank/DDBJ databases. Cited for: NUCLEOTIDE SEQUENCE. Strain: K10 EMBL AAT95422.1.
- 23 Motlagh A, Bhunia AK, Szostek F, Hansen TR, John Son MC, Ray B. 1992. Nucleotide and amino acid sequence of *pap*-gene (pediocin AcH production) in *Pediococcus acidilactici* H. *Letters in Applied Microbiology* 15:45-48.
- 24 Motlagh A, Bukhtiyarova M, Ray B. 1994. Complete nucleotide sequence of pSMB 74, a plasmid encoding the production of pediocin AcH in *Pediococcus acidilactici*. *Letters in Applied Microbiology* 18:305-312.
- 25 Naidu A.S. 2000. *Natural Food Antimicrobial System.* Publisher: CRC Press. p. 542.
- 26 Nissen-Meyer J, Nes IF. 1997. Ribosomally synthesized antimicrobial peptides: their function, biogenesis and mechanism of action. *Arch.Microbiol.* 167: 67-77.
- 27 Nugroho DA, Rahayu, ES. 2003. Ekstraksi dan Karakterisasi Bacteriosin yang dihasilkan oleh *Leuconostoc mesenteroides* SM-22. *Jurnal Teknologi dan Industri. Pangan, Vol. XIV (3):* 214-218.
- 28 Laoye OA, Dodd CER. 2008. Detection and nucleotide sequencing of pediocin in lactic acid bacteria. Submitted (APR-2008) to the EMBL/GenBank/DDBJ databases. Cited for: NUCLEOTIDE SEQUENCE. Strain: PED 01 EMBL ACD45084.1
- 29 Osmanagaoglu O, Kiran F, Nes IF. 2011. A probiotic bacterium, *Pediococcus pentosaceus* OZF, isolated from human breast milk produces pediocin AcH/PA-1. *African Journal of Biotechnology.* 10(11):2070-2079.
- 30 Osmanagaoglu O, Gunduz U, Beyatli Y, Cokmus C. 1998. Purification and Characterization of Pediocin F, A Bacteriocin Produced By *Pediococcus acidilactici* F. *Tr. J. of Biology* 22: 217-228.
- 31 Osmanagaoglu O, Beyatli Y, Ufuk GNDZ. 2001. Isolation and Characterization of Pediocin Producing *Pediococcus pentosaceus* Pep1 from Vacuum-Packed Sausages. *Turk J Biol* 25:133-143.
- 32 Papagianni M, Anastasiadou S. 2009. Pediocins: The bacteriocins of *Pediococci*. Sources, production, properties and applications. *Microbial Cell Factories* 8:3doi:10.1186/1475-2859-8-3.
- 33 Parada, Luis J, Carolina LC, Bianchi APR, Socco CR. 2007. Bacteriocins from Lactic Acid Bacteria: Purification, Properties and use as Biopreservatives. *Brazilian Archives for Biology and Technology* 50: 521 – 542.
- 34 Sambrook J, Fritsh EF, Maniatis T. 1989. *Molecular cloning: A laboratory manual*, 2nd ed. Cold Spring Harbor Laboratory Press, New York.
- 35 Schved F, Lalazar Y, Henis Y, Juven BJ. 1993. Purification, partial characterization and plasmid-linkage of pediocin SJ-1, a bacteriocin produced by *Pediococcus acidilactici*. *J Appl Bacteriol* 74:67-77.
- 36 Todorov SD, Dicks LMT. 2009. Bacteriocin production by *Pediococcus pentosaceus* isolated from marula (*Scerocarya birrea*), *International Journal of Food Microbiology* doi: 10.1016/j.ijfoodmicro.
- 37 Wu CW, Yin LJ, Jiang ST. 2004. Purification and characterization of bacteriocin from *Pediococcus pentosaceus* ACCEL. *Applied and Environmental Microbiology* 52:1146-1151.
- 38 Yang R, Johnson MC, Ray, B. 1992. Novel method to extract large amount of bacteriocin from lactic acid bacteria. *Applied and Environmental Microbiology* 58:3355-3359.